Supporting Information for:

Cyclodidemniserinol trisulfate, a sulfated serinolipid from the Palauan ascidian *Didemnum* guttatum that inhibits HIV-1 integrase

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- 1. Experimental Section (2 pages).
- 2. ¹H NMR spectrum (500 MHz, DMSO- d_6) of **1**.
- 3. ¹³C NMR spectrum (100 MHz, DMSO-*d*6) of **1**.
- 4. COSY spectrum of **1**.
- 5. HMBC spectrum of **1**.
- 6. HSQC-TOCSY spectrum of **1**.

Experimental Section

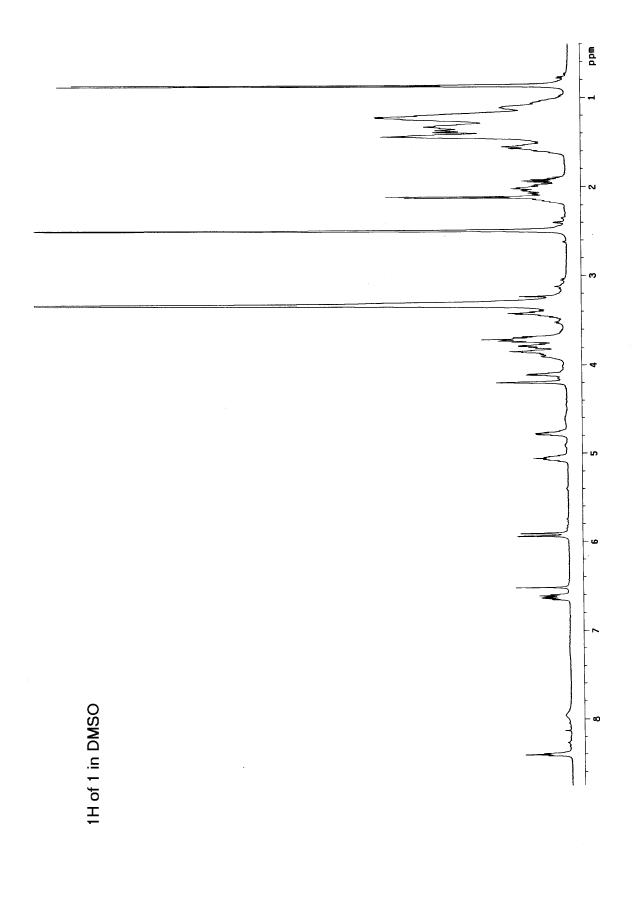
General Methods: The IR and UV spectra were recorded on Perkin-Elmer 1600 and Lambda 3B instruments, respectively. ¹H NMR, COSY, TOCSY, HMQC-TOCSY experiments were recorded on a Varian Inova 300 MHz Spectrometer. The HMBC experiment was acquired on a 500 MHz Varian Gemini spectrometer, and the HETCOR and ¹³C experiments were performed on a 400 MHz Varian spectrometer equipped with a broad band nano probe tuned to ¹³C. Chemical shifts are reported in parts per million, referenced to residual solvent peaks, and coupling constants (*J*), are reported in hertz. High resolution MALDI Fourier transform mass spectra were run on an Ionspec FTMS mass spectrometer at the mass spectrometry facility at the Scripps Research Institute.

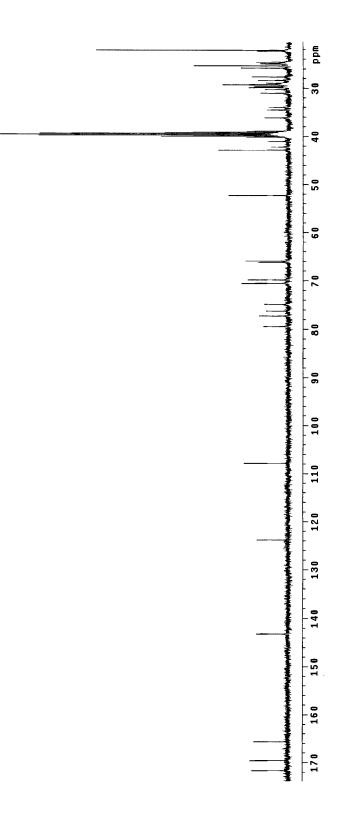
Isolation of cyclodidemniserinol trisulfate (1): The specimen of *Didemnum guttatum* (collection # 96-147) was collected by SCUBA at Ngerchaol Island, Palau and was immediately frozen. The frozen material (400 g wet wt) was extracted with MeOH (4×500 mL), and the extracts were concentrated to obtain a dark green residue (8 g). The residue was partitioned between EtOAc and H₂O, the H₂O-soluble material was lyophilized, and was resuspended in H_2O (10 mL). This solution was loaded onto a C_{18} Sep Pac (12 g), which was subsequently washed with H₂O (150 mL), 90% H₂O/MeOH (100 mL), 75% H₂O/MeOH (100 mL), 50% H₂O/MeOH (100 mL), and 100 % MeOH (200 mL). Activity in the HIV integrase assay was traced to the material eluting in 75% H₂O/MeOH, and this material was further purified by either C_{18} HPLC or C_{18} flash column chromatography. Final purification by HPLC was achieved on a C18 column eluted with a gradient from 90% to 50% H₂O/acetonitrile over 30 minutes to yield cyclodidemniserinol trisulfate (1) as a colorless oil (20 mg, 5×10^{-3} % yield): White solid; $[\alpha]_D$ -26.6° (c 0.073, MeOH); UV (MeOH) λ_{max} 210 nm (ϵ 4500): IR (thin film, polystrene) 2450, 3321, 2068, 1727, 1667, 1625, 1220, 1215, 1119, 1047, 1005 cm⁻¹; ¹H NMR (500 MHz, DMSO d_6) see Table 1); 13C NMR (100 MHz, DMSO- d_6) see Table 1; ESIMS m/z 1039 [M + Na]⁺, 937, 916, 836, 733; HRMS *m/z* 993.2961 [M-Na]⁻, C₃₈H₆₃N₂O₁₉S₃Na₂ requires 933.2983.

Preparation of cyclodidemniserinol (3) for mass spectral analysis:

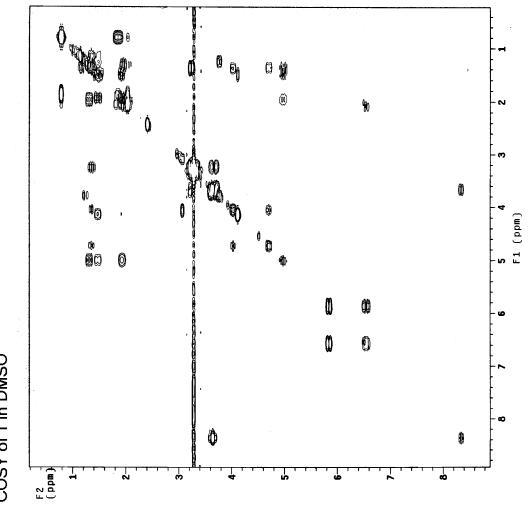
Cyclodidemniserinol trisulfate (1, 0.1 mg) was placed in a thick walled 1 ml vial and dissolved in THF (900 μ L), H₂O (100 μ L), and 1 M H₂SO₄ (5 μ L). The vial was tightly capped and heated to 90° for 1 h, then neutralized by adding solid NaCO₃ (20 mg). The supernatent was removed and dried under reduced pressure. The APCI mass spectra of the resulting residue yielded ions at m/z 733 (M+Na)⁺, 711 (M⁺), and 647 (hydrolysis of isovalerate ester), 376, 334, 203.

Permanganate oxidation of cyclodidemniserinol trisulfate (1): An aqueous solution of cyclodidemniserinol trisulfate (1, 0.75 mg in 600 µL) was adjusted to pH 3 with HCl followed by addition of potssium permanganate (3 mg). The reaction mixture was stirred overnight, partitioned between EtOAc (5 mL) and H₂O (5 mL), and the EtOAc soluble material was dried under reduced pressure. The residue was re-dissolved in CH₂Cl₂ (2 mL) and methylated using diazomethane generated using a DMMNG kit (Aldrich Chemical). The resulting derivative was analyzed using GC-MS on an EC-5 column (Alltech) with a temperature gradient from 50 to 150° over 20 minutes. Standards of dimethyl adipate, dimethyl pimelate, and dimethyl suberate were purchased from Arcos Chemical, and a co-injection experiment determined that dimethyl pimelate co-eluted with a major GC-MS peak observed at 6.9 min in the oxidation product of **1**. The EI spectra of the dimethyl pimelate standard and GC-MS peak observed in the reaction product were identical (*m*/*z* 157 (M-OCH₃, 40%), 128 (43 %), 125 (45%), 115 (98%), 74 (100%).









COSY of 1 in DMSO

